



SYNTHESIS, CHARACTERISATION OF COUMARIN DERIVATIVES AND THEIR ANTI CANCER ACTIVITIES

P. Aravinda Reddy¹, Ramya Sri. S²¹Department of Pharmaceutical analysis, Vaageswari College of Pharmacy, Karimnagar, Telangana-505527, India²Department of Pharmacy, University College of Technology, Osmania University, Hyderabad, Telangana, 500007, India

Received on: 18-12-2016; Revised and Accepted on: 31-12-2016

ABSTRACT

Two sets of coumarins were synthesised, and their IR, ¹H, and ¹³C NMR and mass spectra were all very well characterised. Eight a and eight c, which contain coumarine with an amide-extended chromophore, have shown promise as anti-cancer agents. Our research may lead to more widespread use of coumarines for their potential cancer-fighting properties.

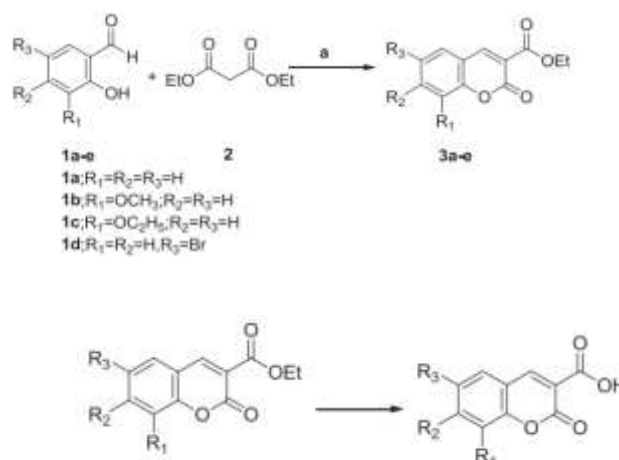
Keywords: Coumarine, Synthesis, Characterisation and anti-cancer.

INTRODUCTION

Cancer is one of the prominent causes of death in the world and based on World Health Organization (WHO) report, more than 13 million cancers death will happen in 2030. It was estimated that one in five people before age 75 will suffer from cancer during their lifetime. Most cancers are recognized by uncontrolled growth of cells without differentiation due to the deregulation of essential enzymes and other proteins controlling cell division and proliferation. Although many efforts for treatment of cancer diseases have been carried out and much progress have been eventuated from diagnosis to treatment of cancer, but some of cancer patients do not respond to therapy or recurrence subsequent initial response. Nevertheless, chemotherapy is a basic approach for the treatment of cancer diseases. One of the most important obstacles in chemotherapy is drug resistance to many anticancer agents. Drug-induced toxicities followed by administering high doses of chemotherapeutic agents to overcome drug resistance were arisen. Accordingly, the discovery of new anticancer agent with promising activity and high therapeutic index is an urgent need. Coumarin (1, 2H-chromen-2-one or 2H-1-benzopyran-2-one) nucleus is a bicyclic heterocycle consisting of benzene and 2- pyrone rings (Fig. 1). Coumarins, a part of flavonoid group of plant secondary metabolite, are a wide class of natural and synthetic compounds that showed versatile pharmacological activities including anti-inflammatory, antioxidant, antinociceptive, hepatoprotective,

antithrombotic, antiviral, antimicrobial, antituberculosis, anti-carcinogenic, antidepressant, antihyperlipidemic and anticholinesterase activities. Several natural and synthetic drugs containing coumarin scaffold are clinically well-known agents. For example, hymecromone (4-methylumbelliferone, 2) was used as choleric and antispasmodic agent. Scopoletin (6-methoxy-7- hydroxycoumarin, 3) has been isolated from several plant species and has antioxidant, hepatoprotective, anti-inflammatory and antifungal properties. Carbochromen (4) has beneficial effect in coronary disease. 4-Hydroxycoumarin derivatives acenocoumarol (5), phenprocoumon (6), warfarin (7), difenacoum (8), and brodifacoum (9) are anticoagulant agents that act as vitamin K antagonists.

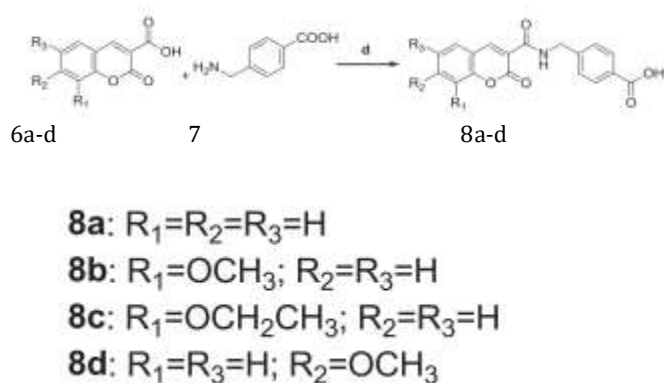
Synthetic Scheme:



*Corresponding author:

P. Aravinda Reddy
Department of Pharmaceutical analysis
Vaageswari College of Pharmacy, Karimnagar
Telangana-505527
India

Email: surapharmalabs1@gmail.comDOI: <https://doi.org/10.5281/zenodo.7701497>



Scheme 1. Reagents and conditions: (a) Piperidine, EtOH, reflux; (b) NaOH (2 N), rt and then HCl (2 N); (c) CDI, THF, reflux; (ii) TFA, rt.

- Infrared spectra are recorded on Perkin Elmer model 283B and Nicolet 740 FT-IR. Instruments and values are given in cm⁻¹
- Proton Nuclear Magnetic Resonance spectra are recorded on Varian Gemini-200, Varian Unity-200 and Avance-400 MHz Bruker UX-NMR instrument. The samples are made in chloroform-d(1:1) or/and DMSO-d₆ using tetramethylsilane (MeSi) as the internal standard and are given in the δ scale.
- ESI Mass spectra were recorded on a Micro mass Quattro LC using ESI+ software with capillary voltage 3.98 kV and ESI mode positive ion trap detector.
- High-resolution mass spectra (HRMS) were recorded on a QSTAR XL Hybrid MS-MS mass spectrometer.
- Elemental analysis is carried out on VARIO EL, se Elementor.
- Analytical Thin-layer Chromatography (TLC) is performed on pre-coated silica-gel-60 F
- All extracts are extracted with ethyl acetoacetate and water and concentrated at reduced pressure on Buchi-R-3000 rotary evaporator below 50°C. Yields reported are isolated yields of materials judged homogenous by TLC and NMR spectroscopy.
- Employing TLC techniques using appropriate solvent system for development monitored all the reasons. Moisture sensitive reactions are carried out by standard syringe-septum techniques. Dry ether, dry toluene is made by distilling them from sodium benzophenone ketyl and dry methanol is prepared by using potassium hydroxide.
- 254(0.5mm) glass plates. Visualisation of the spots on TLC plates is achieved either by iodine vapour or UV light.
- Melting Points were recorded on Melter Fp-51 instrument and were uncorrected.

Experimental Procedure:

All reagents and solvents were purchased from Merck & Co., Inc. (Darmstadt, Germany) and Acros Organics (ThermoFisher, Belgium) and used without further purification. Entinostat and vorinostat were purchased from EuroAsia Chemicals (@www.euroasiarnd.com). The progress of all reactions was monitored by thin-layer chromatography with 0.25 mm Silica gel plates (60 GF-254-Merck & Co (Darmstadt, Germany)) and

visualized using UV light and iodine vapor. Melting points were measured with an electrothermal melting point apparatus (Stafford, UK) and were uncorrected. Infrared spectra were recorded on a Perkin Elmer Model 1420 spectrometer (KBr disks, Massachusetts, USA). ¹H NMR and ¹³C NMR spectra were determined at 300 MHz and 75 MHz, respectively (DMSO-d₆, TMS) on a Bruker FT-300 MHz instrument (Karlsruhe, Germany). The chemical shifts (δ) and coupling constants (J) are expressed in parts per million and Hertz, respectively. Spin multiples are given as s (singlet), d (doublet), t (triplet), q (quartet), s (sextet), m (multiplet). Mass spectra were obtained from a 6410 Agilent LC-MS triple quadrupole mass spectrometer (LC-MS, Santa Clara, USA) with an electrospray ionization (ESI) interface. Elemental analyses were performed on a Cos-Tec model EAS 4010 instrument (Cernusco, Italy) and the results are within ±0.4% of the theoretical values. Cell lines, including HCT116, A2780, MCF7, A549, HL60, PC3 and Huvec purchased from Pasteur Institute Cell Bank of IRAN (Tehran, IRAN).

General procedure for the synthesis of ethyl 2-oxo-2H-chromene-3-carboxylates (3a-d)

Diethyl malonate (2.11 mmol) and piperidine (2 mL) were added to a solution of the appropriate salicylaldehyde derivatives (1a-d) (10 mmol) in ethanol (20 mL) and the resulting mixture was refluxed for 14–15 h until the disappearance of the starting materials (monitored by TLC). After cooling, the crude product was filtrated, washed with cold ethanol and recrystallized from ethanol.

General procedure for the synthesis of 2-oxo-2H-chromene-3-carboxylic acids (4a-d)

Esters (3a-v) were dissolved in aqueous sodium hydroxide (2 N, 20 mL) and were stirred at rt for 5–7 h. After cooling, solution was acidified with hydrochloric acid (2 N) until a white precipitate formed. The white solid was filtrated, washed with water and dried to yield acids (4a-d).

General procedure for the synthesis of 4-((2-oxo-2H-chromene-3-carboxamido) methyl) benzoic acid derivatives (6a-d) the corresponding 2-oxo-2H-chromene-3-carboxylic acids (3a-d, 5 mmol) was added to a suspension of 1, 1'-carbonyldimidazole (CDI, 25 mmol) in dry tetrahydrofuran (THF, 10 mL) and the mixture was stirred for 2 h at rt. Then 4-(aminomethyl) benzoic acid (5, 25 mmol) and trifluoroacetic acid (TFA, 1.2 mL) were added and stirred for additional 10 h at rt. The mixture was evaporated to remove THF and extracted with EtOAc, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. Further purification was done with preparative thin layer chromatography.

4-((2-oxo-2H-chromene-3-carboxamido) methyl) benzoic acid (6a) White solid; yield: 78%; mp: 261–263 °C; R_f = 0.56 (petroleum ether: ethyl acetate = 3:1); IR (KBr): ν_{max} (cm⁻¹) 3312 (NH), 1709 1602 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 4.63 (d, 2H, J = 6.0 Hz, CH₂), 7.44–7.54 (m, 4H, Ar-H), 7.77 (t, 1H, J = 6.0 Hz, Ar-H), 7.92 (d, 2H, J = 9.0 Hz, Ar-H), 7.99 (d, 1H, J = 9.0 Hz, Ar-H), 8.90 (s, 1H, CH), 9.23 (t, 1H, J = 6.0 Hz, NH); ¹³C NMR (75 MHz, DMSO-d₆) δ 43.04, 116.62, 118.94, 119.57, 125.62, 127.78, 129.88, 130.06, 130.75, 134.61, 144.56, 148.08, 154.40, 160.73, 161.96, 167.60; Anal. calcd. for C₁₈H₁₃N₂O₅: C, 66.87; H, 4.05; N, 4.33; found: C 66.80, H 4.09, N 4.24.

4-((8-methoxy-2-oxo-2H-chromene-3-carboxamido) methyl) benzoic acid (6b) White solid; yield: 73%; mp: 238–240 °C; Rf = 0.49 (petroleum ether: ethyl acetate = 3:1); IR (KBr): ν_{max} (cm⁻¹) 3347 (NH), 1712 & 1606 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 3.95 (s, 1H, OCH₃), 4.62 (d, 2H, J = 6.0 Hz, CH₂), 7.35–7.54 (m, 5H, Ar-H), 7.91 (d, 1H, J = 8.1 Hz, Ar-H), 8.85 (s, 1H, CH), 9.24 (t, 1H, J = 5.7 Hz, NH); ¹³C NMR (75 MHz, DMSO-d₆) δ 43.05, 56.65, 116.53, 119.46, 119.57, 121.62, 125.54, 127.81, 129.36, 137.81, 143.69, 144.56, 146.73, 148.30, 160.44, 161.83, 167.60; Anal. calcd. for C₁₉H₁₅N₂O₆: C, 64.59; H, 4.28; N, 3.96; found: C 64.66, H 4.20, N 4.05.

4-((8-ethoxy-2-oxo-2H-chromene-3-carboxamido) methyl) benzoic acid (6c) Pale yellow solid; yield: 79%; mp: 246–248 °C; Rf = 0.43 (petroleum ether: ethyl acetate = 3:1); IR (KBr): ν_{max} (cm⁻¹) 3335 (NH), 1707 and 1653 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 1.42 (t, 3H, J = 6.9 Hz, CH₃), 4.20 (q, 2H, J = 6.9 Hz, OCH₂), 4.62 (d, 2H, J = 6.0 Hz, CH₂), 7.34 (d, 1H, J = 7.5 Hz, Ar-H), 7.42–7.48 (m, 3H, Ar-H), 7.52 (dd, 1H, J₁ = 7.5 Hz, J₂ = 1.2 Hz, Ar-H), 7.92 (d, 2H, J = 8.1 Hz, Ar-H), 8.86 (s, 1H, CH), 9.23 (t, 1H, J = 6.0 Hz, NH); ¹³C NMR (75 MHz, DMSO-d₆) δ 15.04, 43.03, 65.02, 117.43, 119.56, 119.59, 121.62, 125.56, 127.82, 129.89, 137.82, 143.79, 144.63, 145.98, 148.37, 160.53, 161.96, 167.66; Anal. calcd. for C₂₀H₁₇N₂O₆: C, 65.39; H, 4.66; N, 3.81; found: C 65.30, H 4.73, N 3.75.

4-((7-methoxy-2-oxo-2H-chromene-3-carboxamido) methyl) benzoic acid (6d) Light brown; yield: 89%; mp: 242–244 °C; Rf = 0.46 (petroleum ether: ethyl acetate = 3:1); IR (KBr): ν_{max} (cm⁻¹) 3346 (NH), 1701, 1659 & 1614 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 3.89 (s, 3H, OCH₃), 4.62 (d, 2H, J = 6.0 Hz, CH₂), 7.02 (dd, 1H, J₁ = 8.7 Hz, J₂ = 2.4 Hz, Ar-H), 7.09 (d, 1H, J = 2.1 Hz, Ar-H), 7.45 (d, 2H, J = 8.1 Hz, Ar-H), 7.90 (m, 3H, Ar-H), 8.83 (s, 1H, CH), 9.17 (t, 1H, J = 6.0 Hz, NH), 12.91 (brs, 1H, OH); ¹³C NMR (75 MHz, DMSO-d₆) δ 42.98, 56.67, 100.69, 112.54, 114.05, 115.16, 127.77, 129.84, 129.90, 131.99, 144.77, 148.47, 156.64, 161.24, 162.20, 164.90, 167.65; Anal. calcd. for C₁₉H₁₅N₂O₆: C, 64.59; H, 4.28; N, 3.96; found: C 64.48, H 4.34, N 3.89.

N-(4-((2-aminophenyl) carbamoyl) benzyl)-2-oxo-2H-chromene-3-carboxamide (8a) White solid; yield: 90%; mp: 207–209 °C; Rf = 0.76 (petroleum ether: ethyl acetate = 3:2); IR (KBr): ν_{max} (cm⁻¹) 3404 and 3322 (NH and NH₂), 1720, 1653 and 1609 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 4.64 (d, 2H, J = 6.0 Hz, CH₂), 4.90 (s, 2H, NH₂, D₂O exchangeable), 6.62 (t, 1H, J = 6.6 Hz, Ar-H), 6.79 (d, 1H, J = 7.5 Hz, Ar-H), 6.93 (t, 1H, J = 7.5 Hz, Ar-H), 7.18 (d, 1H, J = 6.0 Hz, Ar-H), 7.48–7.55 (m, 4H, Ar-H), 7.75 (t, 1H, J = 6.0 Hz, Ar-H), 7.98 (m, 2H, Ar-H), 8.90 (s, 1H, CH), 9.22 (t, 1H, J = 6.0 Hz, NH, D₂O exchangeable), 9.64 (s, 1H, NH, D₂O exchangeable); ¹³C NMR (75 MHz, DMSO-d₆) δ 43.03, 116.63, 116.79, 118.95, 119.58, 123.70, 125.63, 126.93, 127.05, 127.63, 128.36, 129.93, 130.75, 133.75, 134.62, 143.00, 148.05, 154.39, 160.78, 161.91, 165.53; LCMS(ESI, m/z): (M+1)⁺ 414.0, [2 M + 1]⁺ 827.2; Anal. calcd. for C₂₄H₁₉N₃O₄: C, 69.72; H, 4.63; N, 10.16; found: C 69.78, H 4.69, N 10.06.

N-(4-((2-aminophenyl) carbamoyl) benzyl)-8-methoxy-2-oxo-2H-chromene-3-carboxamide (8b)

White solid; yield: 80%; mp: 213–215 °C; Rf = 0.65 (petroleum ether: ethyl acetate = 3:2); IR (KBr): ν_{max} (cm⁻¹) 3402 and 3342 (NH and NH₂), 1701, 1659 and 1601 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 3.95 (s, 3H, OCH₃), 4.58 (d, 2H, J = 6.0 Hz, CH₂), 4.95 (s, 2H, NH₂), 6.64 (t, 1H, J = 6.0 Hz, Ar-H), 6.79 (d, 1H, J = 6.6 Hz, Ar-H), 6.97 (t, 1H, J = 6.6 Hz, Ar-H), 7.15 (d, 1H, J = 6.6

Hz, Ar-H), 7.30–7.50 (m, 5H, Ar-H), 7.95 (d, 2H, J = 6.6 Hz, Ar-H), 8.87 (s, 1H, CH), 9.23 (t, 2H, J = 6.0 Hz, NH), 9.66 (s, 1H, NH); ¹³C NMR (75 MHz, DMSO-d₆) δ 43.04, 56.69, 116.59, 116.80, 119.51, 119.70, 121.66, 123.71, 125.58, 126.93, 127.04, 127.66, 128.36, 133.76, 142.99, 143.41, 143.72, 146.77, 148.27, 160.49, 161.88, 161.93, 165.59; LCMS(ESI, m/z): [M+1]⁺ 444.0, [2 M + 1]⁺ 887.3; Anal. calcd. for C₂₅H₂₁N₃O₅: C, 67.71; H, 4.77; N, 9.48; found: C 67.82, H 4.82, N 9.56.

N-(4-((2-aminophenyl) carbamoyl) benzyl)-8-ethoxy-2-oxo-2H-chromene-3-carboxamide (8c) White solid; yield: 87%; mp: 218–220 °C; Rf = 0.46 (petroleum ether: ethyl acetate = 3:2); IR (KBr): ν_{max} (cm⁻¹) 3400 and 3326 (NH and NH₂), 1720, 1653 and 1613 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 1.43 (t, 3H, J = 6.9 Hz, CH₃), 4.23 (q, 2H, J = 6.9 Hz, OCH₂), 4.63 (s, 2H, J = 5.4 Hz, CH₂), 4.92 (s, 2H, NH₂), 6.61 (t, 1H, J = 6.9 Hz, Ar-H), 6.78 (d, 1H, J = 6.9 Hz, Ar-H), 7.00 (t, 1H, J = 6.9 Hz, Ar-H), 7.17 (d, 1H, J = 7.2 Hz, Ar-H), 7.33–7.54 (m, 5H, Ar-H), 7.95 (d, 2H, J = 8.1 Hz, Ar-H), 8.87 (s, 1H, CH), 9.24 (t, 1H, J = 6.0 Hz, NH), 9.66 (s, 1H, NH); ¹³C NMR (75 MHz, DMSO-d₆) δ 15.04, 43.01, 65.00, 116.56, 116.74, 117.37, 119.60, 121.61, 123.65, 123.76, 125.58, 126.95, 127.15, 127.65, 128.36, 133.76, 143.00, 143.50, 143.76, 145.98, 148.35, 160.57, 161.90, 165.61; LCMS(ESI, m/z): 458.0 [M+1]⁺, 915.1 [2 M + 1]⁺; Anal. calcd. for C₂₆H₂₃N₃O₅: C, 68.41; H, 5.07; N, 9.19; found: C 68.29, H 5.01, N 9.05.

N-(4-((2-aminophenyl) carbamoyl) benzyl)-7-methoxy-2-oxo-2H-chromene-3-carboxamide (8d) Yellow solid; yield: 92%; mp: 212–214 °C; Rf = 0.53 (petroleum ether: ethyl acetate = 3:2); IR (KBr): ν_{max} (cm⁻¹) 3402 and 3342 (NH and NH₂), 1701, 1659 and 1601 (CO); ¹H NMR (300 MHz, DMSO-d₆) δ 3.91 (s, 3H, OCH₃), 4.62 (d, 2H, J = 5.4 Hz, CH₂), 4.94 (brs, 2H, NH₂), 6.61 (t, 1H, J = 7.2 Hz, Ar-H), 6.78 (d, 1H, J = 7.8 Hz, Ar-H), 7.00 (t, 1H, J = 7.2 Hz, Ar-H), 7.05 (m, 3H, Ar-H), 7.48 (d, 2H, J = 8.4 Hz, Ar-H), 7.95 (m, 3H, Ar-H), 8.87 (s, 1H, CH), 9.19 (t, 1H, J = 5.4 Hz, NH), 9.66 (s, 1H, NH); ¹³C NMR (75 MHz, DMSO-d₆) δ 42.85, 56.75, 100.78, 112.62, 114.14, 115.32, 116.59, 116.79, 123.68, 126.95, 127.07, 127.61, 128.36, 132.07, 133.70, 143.12, 143.42, 148.48, 156.70, 161.26, 162.19, 164.94, 165.53; LCMS(ESI, m/z): 444.0 [M+1]⁺, 887.3 [2 M + 1]⁺; Anal. calcd. for C₂₅H₂₁N₃O₅: C, 67.71; H, 4.77; N, 9.48; found: C 67.86, H 4.63, N 9.59.

Biological Activity

Anticancer activity has been carried out for the synthesized compounds using MDAMB (breast cancer) cell line by MTT Assay. Cell proliferation and viability was determined by 3-[4, 5-dimethylthiazol-2-yl]-2, 5-diphenyltetrazolium bromide (MTT) assay [60]. The pale yellow colored tetrazolium salt (MTT) reduces to a dark blue water-insoluble formazan by metabolically active cells and that is measured quantitatively after soluble in DMSO. The absorbance of the soluble formazan is directly proportional to the number of viable cells. Cells were seeded at a density of 1×10⁴ cells in 200 μ L of medium per well of 96-well plate. The 96-well microtiter plates were incubated for 24 h prior to addition of the experimental compounds. Cells were treated with vehicle alone (0.4% DMSO) or compounds (drugs were dissolved in DMSO previously) at different concentrations (1, 10 and 25 μ M) of test compounds for 48 hours. The assay was completed with the addition of MTT (5 %, 10 μ L) and incubated for 60 min at 37°C. The supernatant was aspirated and plates were air dried and the MTT-formazan crystals dissolved in 100 μ L of DMSO. The optical density (O.D.) was measured at 560 nm using TECAN multimode reader. The

growth percentage of each treated well of 96 well plate have been calculated based on test wells relative to control wells [61].

The cell growth inhibition was calculated by generating dose response curves as a plot of the percentage of surviving cells versus drug concentration. Anti-proliferative activity of the cancer cells to the test compounds was expressed in terms of IC₅₀ value, which defines as a concentration of compound that produced 50% absorbance reduction relative to control.

Results:

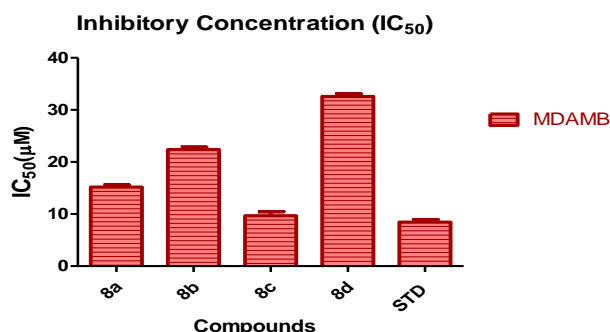
The compounds 8a and 8c showed strong potent activity with IC₅₀ values around 9.68 and 15.2 µg/ml against MDAMB cell lines where as 8b and 8d were showed poor inhibition activity.

Table 5:IC₅₀ (µM) for the synthesized compounds and (STD) on the cells MDAMB (breast cancer) determined by MTT assay:

Compound code	MDAMB (breast cancer)
8a	15.2 ± 0.39
8b	22.39 ± 0.47
8c	9.68 ± 0.78
8d	32.57 ± 0.53
STD	8.45 ± 0.49

STD= Fluorouracil

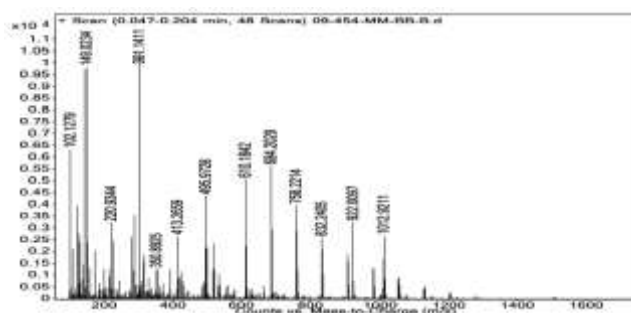
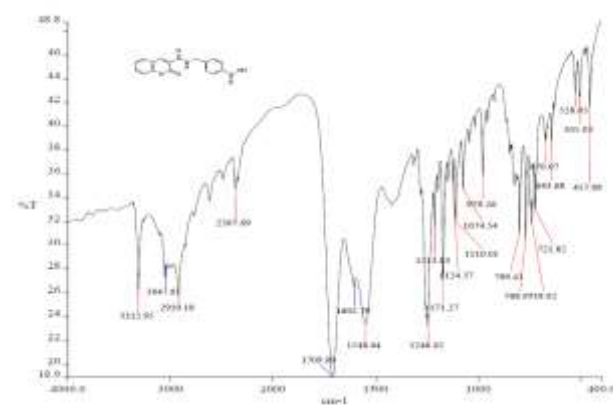
Data represented as mean ± standard deviation

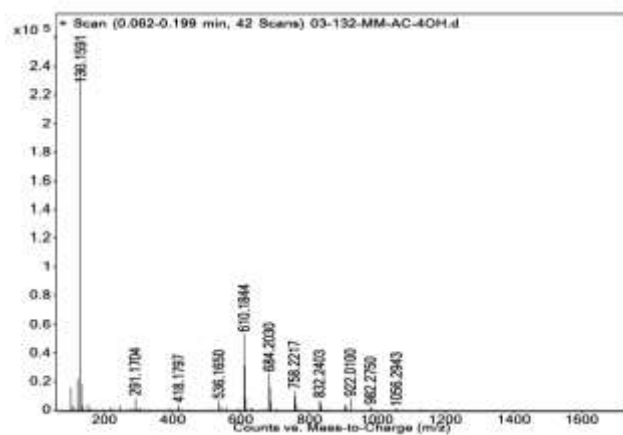
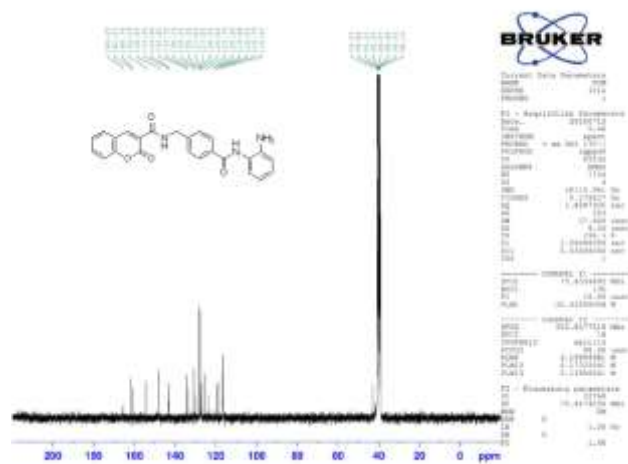
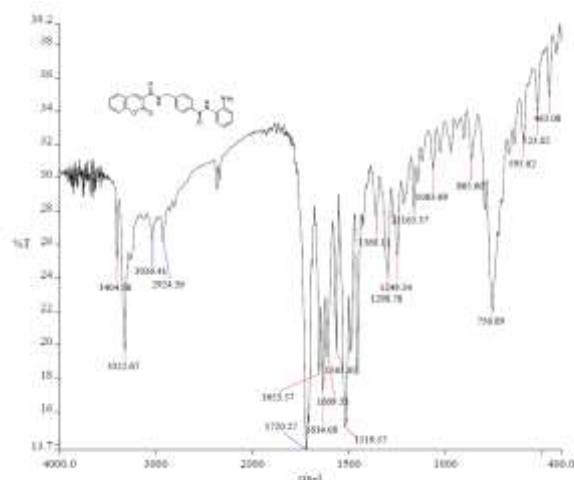


Graph 1: Comparison of IC₅₀ (µM) for the synthesized compounds and (STD) on the MDAMB cells

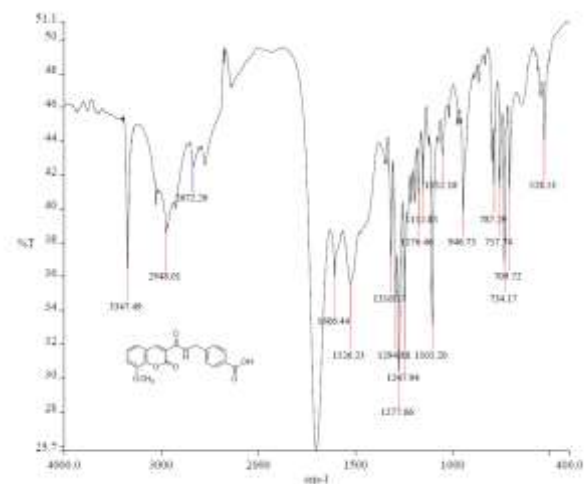
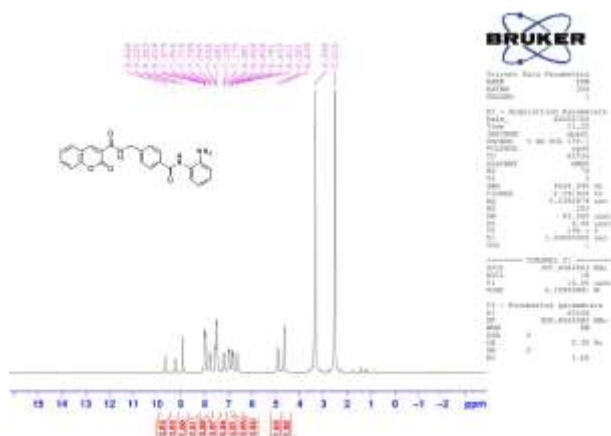
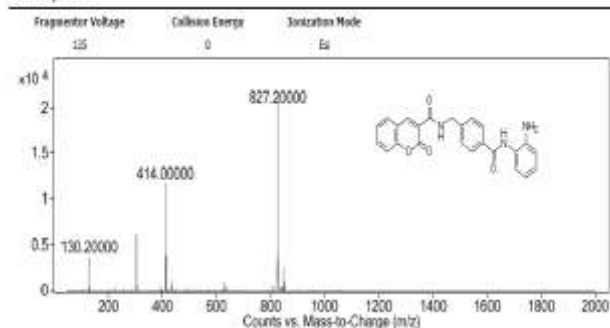
Conclusion

Thus the coumarin derivatives 8a and 8c serve as good leads for further studies to develop potent cytotoxic agents.





User Spectra



CONCLUSION

An effective review of literature had been done on coumarines; synthesized two sets of coumarines and well characterize by IR, ¹H & ¹³C NMR, Mass Spectra. 8a and 8c were found to be potential towards anti-cancer activity because of coumarine with an extended chromophore by amide linkage. We hope our study would be an expansion of anti-cancer activity of coumarines.

ACKNOWLEDGEMENT

The Authors are thankful to the Management and Principal, Department of Chemistry, Vaageswari College of Pharmacy, Karimnagar, for extending support to carry out the research work. Finally, the authors express their gratitude to the Sura Labs, Dilsukhnagar, Hyderabad, for providing research equipment and facilities.

REFERENCES:

- [1] A. Jemal, F. Bray, M.M. Center, J. Ferlay, E. Ward, D. Forman, *CA Cancer J. Clin.* 61 (2011) 69e90.
- [2] S.-K. Kim, *Handbook of Anticancer Drugs from Marine Origin*, Springer International Publishing, Switzerland, 2015.
- [3] J. Ferlay, H.R. Shin, F. Bray, D. Forman, C. Mathers, D.M. Parkin, *GLOBOCAN 2008 v1.2. Cancer Incidence and Mortality Worldwide: IARC Cancer Base No. 10*, International Agency for Research on Cancer, Lyon, France, 2010.
- [4] M. Mareel, A. Leroy, *Physiol. Rev.* 83 (2003) 337e376.
- [5] J. Wesche, K. Haglund, E.M. Haugsten, *Biochem. J.* 437 (2011) 199e213.
- [6] S.K. Grant, *Cell. Mol. Life Sci.* 66 (2009) 1163e1177.
- [7] G.I. Solyanik, *Exp. Oncol.* 32 (2011) 181e185.
- [8] S. Vijayaraghavalu, C. Peetla, S. Lu, V. Labhasetwar, *Mol. Pharm.* 9 (2012) 2730e2742.
- [9] S. Riedl, D. Zweytick, K. Lohner, *Chem. Phys. Lipids* 164 (2011) 766e781.
- [10] K.C. Fylaktakidou, D.J. Hadjipavlou-Litina, K.E. Litinas, D.N. Nicolaides, *Curr. Pharm. Des.* 10 (2004) 3813e3833.
- [11] (a) I. Kostova, S. Bhatia, P. Grigorov, S. Balkansky, V.S. Parmar, A.K. Prasad, L. Saso, *Curr. Med. Chem.* 18 (2011) 3929e3951;
- (b) M. Khoobi, S. Emami, G. Dehghan, A. Foroumadi, A. Ramazani, A. Shafiee, *Arch. Pharm. Chem. Life Sci.* 344 (2011) 588e594.
- [12] (a) T.A.A. Barros, L.A.R. Freitas, J.M.B. Filho, X.P. Nunes, A.M. Giulietti, G.E. Souza, R.R. Santos, M.B.P. Soares, C.F. Villarreal, *J. Pharm. Pharmacol.* 62 (2010) 205e213; (b) M. Alipour, M. Khoobi, S. Emami, S. Fallah-benakohal, S.F. Ghasemi-Niri, M. Abdollahi, A. Foroumadi, A. Shafiee, *Daru J. Pharm. Sci.* 22 (2014) 9.
- [13] M. Atmaca, H.M. Bilgin, B.D. Obay, H. Diken, M. Kelle, E. Kale, *J. Physiol. Biochem.* 67 (2011) 569e576.
- [14] X.-M. Peng, G.L. Damu, C. Zhou, *Curr. Pharm. Des.* 19 (2013) 3884e3930.
- [15] M. Curini, F. Epifano, F. Maltese, M.C. Marcotullio, S.P. Gonzales, J.C. Rodriguez, *Aust. J. Chem.* 56 (2003) 59e60.
- [16] (a) D.A. Ostrov, J.A. Hernandez Prada, P.E. Corsino, K.A. Finton, N. Le, T.C. Rowe, *Antimicrob. Agents Chemother.* 51 (2007) 3688e3698; (b) S. Emami, A. Foroumadi, M.A. Faramarzi, N. Samadi, *Arch. Pharm. Chem. Life Sci.* 341 (2008) 42e48.
- [17] N.A. Gormley, G. Orphanides, A. Meyer, P.M. Cullis, A. Maxwell, *Biochemistry* 35 (1996) 5083e5092.
- [18] A. Manvar, A. Bavishi, A. Radadiya, J. Patel, V. Vora, N. Dodia, K. Rawal, A. Shah, *Bioorg. Med. Chem. Lett.* 21 (2011) 4728e4731.
- [19] R.V. Nair, E.P. Fisher, S.H. Safe, C. Cortez, R.G. Harvey, J. DiGiovanni, *Carcinogenesis* 12 (1991) 65e69.
- [20] K.V. Sashidhara, A. Kumar, M. Chatterjee, K.B. Rao, S. Singh, A.K. Verma, G. Palit, *Bioorg. Med. Chem. Lett.* 21 (2011) 1937e1941.
- [21] B. Yuce, O. Danis, A. Ogan, G. Sener, M. Bulut, A. Yarat, *Arzneim. Forsch. Drug Res.* 59 (2009) 129e134.
- [22] (a) S.F. Razavi, M. Khoobi, H. Nadri, A. Sakhteman, A. Moradi, S. Emami, A. Foroumadi, A. Shafiee, *Eur. J. Med. Chem.* 64 (2013) 252e259;
- (b) A. Asadipour, M. Alipour, M. Jafari, M. Khoobi, S. Emami, H. Nadri, A. Sakhteman, A. Moradi, V. Sheibani, F. Homayouni Moghadam, A. Shafiee, A. Foroumadi, *Eur. J. Med. Chem.* 70 (2013) 623e630;
- (c) M. Alipour, M. Khoobi, A. Moradi, H. Nadri, F. Homayouni Moghadam, S. Emami, Z. Hasanpour, A. Foroumadi, A. Shafiee, *Eur. J. Med. Chem.* 82 (2014) 536e544;
- (d) S. Mohammad Bagheri, M. Khoobi, H. Nadri, A. Moradi, S. Emami, L. Jalili Baleh, F. Jafarpour, F. Homayouni Moghadam, A. Foroumadi, A. Shafiee, *Chem. Biol. Drug Des.* (22 May 2015),
- [23] A. Abate, V. Dimartino, P. Spina, P.L. Costa, C. Lombardo, A. Santini, M. Del Piano, P. Alimonti, *Drugs Exp. Clin. Res.* 27 (2001) 223e231.

How to cite this article:

P. Aravinda Reddy. et al., SYNTHESIS, CHARACTERISATION OF COUMARIN DERIVATIVES AND THEIR ANTI CANCER ACTIVITIES. J Pharm Res, 2016; 05(12): 282- 288 DOI: <https://doi.org/10.5281/zenodo.7701497>

Conflict of interest: The authors have declared that no conflict of interest exists.

Source of support: Nil